Protein information sheet

Gene name	**PFN1**
Uniprot ID	P07737
Region	M1 – Y140

Description	profilin 1
Synonyms	ALS18
Construct ID	PFN1A-c000
Protein sequence	*SM*AGWNAYIDNLMADGTCQDAAIVGYKDSPSVWAAVPGKTFVNITPAEVGVLV GKDRSSFYVNGLTLGGQKCSVIRDSLLQDGEFSMDLRTKSTGGAPTFNVTVTK TDKTLVLLMGKEGVHGGLINKKCYEMASHLRRSQY*SSKGGYGLNDIFEAQKIE WHE*

Protein mass	**with biotin, + 226.1**	**Protein pI**	**Extinction (M^{-1} cm^{-1})**
17532	17758	6.79	25440

Expression host	E. coli BL21(DE3)-R3-BirA
Expression medium	Terrific broth (TB) with 50 µg/ml kanamycin, 50 µg/ml Streptomycin and Chloramphenicol (34 µg/ml) included in starter cultures; chloramphenicol omitted in final growth. Biotin (10 mM stock): dissolve 24mg of d-biotin in 10 ml of warm (microwaved) 10mM bicine buffer (pH 8.3) and sterilize of the solution with a syringe and a 0.2 micron filter.
Transformation and storage	Cells were transformed with the expression plasmid and selected on LB-agar plates containing kanamycin, streptomycin and chloramphenicol. Several colonies were picked from the overnight transformation and used to inoculate cultures in LB supplemented with KAN/Cam/Str which were grown overnight and stored as glycerol (15% v/v) stock at -80°C.
Expression and *in vivo* biotinylation	10ml starter cultures in LB supplemented with KAN/Cam/Str were grown overnight at 37°C with shaking at 250rpm. Next day, 10 ml was used to inoculate 1L of TB in a 2LUltraYield flask supplemented with Streptomycin 50µg/ml and Kanamycin 50µg/ml. **Note:** the medium need not contain chloramphenicol at this step; the pRARE2 plasmid is not lost during the growth process and also ensures a more predictable growth. Seal flask with an adhesive air porous film during cell growth. Culture was grown in a 37°C incubator with shaking at 200rpm. Cell growth was monitored until mid-log phase was reached and cell density OD_{600nm} was approximately 2.00 – 3.00 in TB. Incubator temperature was lowered to 18ºC and culture flask allowed to cool. After 30 - 45 minutes protein expression was induced by the

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	addition of IPTG at 0.1mM final concentration and Biotin solution (10 ml/ 1L culture). Incubation of the culture was continued for 16 hours at 18°C. Next day, the cells were collected by centrifugation (15 min, 5,000 RPM in JLA-8.100 rotors). The supernatants were discarded and the cell pellets transferred into 50ml conical polypropylene tubes. Cell pellets were weighed, frozen and stored at -80°C.
Purification buffers	2x lysis buffer: 100 mM HEPES, pH 7.5, 1.0 M NaCl, 10% v/v glycerol, 1mM TCEP, 10mM imidazole. Protease inhibitors (1:200 Merck cocktail IV) were added before use. 1x lysis buffer: 50 mM HEPES, pH 7.5, 0.5 M NaCl, 5% v/v glycerol, 1 mM TCEP, 10 mM imidazole Wash buffer: 50 mM HEPES, pH 7.5, 0.5 M NaCl, 5% v/v glycerol, 1 mM TCEP, 40 mM imidazole Elution buffers: 50 mM Hepes pH 7.5, 500mM NaCl, 5% Glycerol, 1mM TCEP, 100mM, 150mM, 250 mM, 500 mM Imidazole, respectively GF buffer: 50 mM HEPES, pH 7.5, 0.5M NaCl, 5% v/v glycerol, 1 mM TCEP PEI 5%, pH 7.5: 5% polyethyleneimine (diluted from 50% stock, Sigma) and **titrated to pH 7.5 with HCl.**
Purification step 1: IMAC	**Cell Lysis:** Frozen cell pellets from -80°C were thawed by intermittently placing in a 37°C water bath, then transferred directly to ice. Cells were re-suspended in pre-chilled lysis buffer by vortex or pipetting gentle until cells were completely in suspension. 1 volume (v/w) of 2x lysis buffer was added, and the cells were thoroughly suspended. After further addition of 3 volumes (3 ml per 1g of pellet) of 1x lysis buffer the cells were broken by sonication. The lysate was then transferred to a 50ml Beckman polypropylene centrifuge tube. Cell debris and DNA from the lysate was precipitated by adding Polyethylenimine (PEI) to the tube from a 5% stock solution to a final concentration of 0.15% and incubated on ice for 15 – 30 mins for precipitation. Cell lysates were clarified by centrifugation using a JA25.50 rotor at 21,500rpm for 60 min at 4°C. The cleared supernatant was collected and filtered using a 0.45um acrodisc syringe filter and transferred to a fresh 50ml-conical polypropylene tube. **Ni^{2+}-IDA IMAC (column 1):** A 25ml Econo-Pac polypropylene column (BioRad) was packed with 5ml of Ni-Sepharose resin and equilibrated in 1x lysis buffer. The cell lysate was loaded onto the column and passed through resin at gravity flow. The unbound flow-through was collected, a wash step of 10 column volumes of wash buffer was included and collected as the wash fraction. Protein was eluted in four intermediate fractions with a step gradient of 5ml imidazole elution buffers; 100mM, 150mM, 250mM and 500mM imidazole, respectively, each gradient eluted fraction was collected. IMAC fractions were analysed by SDS-PAGE gel for expected molecular weight and purity, the selected eluted fractions were pooled together.

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	Mass spectrometry characterisation: Identity and biotinylation efficiency was verified by mass spectrometry, using a Quadrupole-Time of flight (Q-TOF) analyser, Biotin quantitation is measured by: MW of biotinylated protein = MW of sequenced Protein + 226 Da (biotinylation modification). PFN1 biotinylation efficiency was observed to be incomplete with ~60% of protein in biotinylated form and ~40% un-biotinylated protein.
Step 2: Removal of His_6 tag and Ni^{2+}-IDA reverse binding	**Enzymatic removal of histidine tag:** Separate cleaved protein from any un-cleaved protein by passing over a Ni^{2+}-IDA column. Cleaved protein may be recovered in the flow-through fractions. Non-specific contaminating proteins with any un-cleaved protein bound to the column can be eluted with imidazole. Selected, pooled eluted fractions from Ni^{2+}-IDA IMAC were concentrated using a 10,000 Amicon-Ultra MWCO centrifugal concentrator to approximately 10mls in volume. Cleavage of the histidine tag was performed by the addition of TEV protease in a 1:20 w/w ratio to the pooled eluted protein fractions. The protein sample plus TEV was transferred to Snakeskin dialysis tubing (3,500 MWCO) and dialysed in 1L Gel Filtration buffer overnight at 4°C on a magnetic stirrer plate for digestion. Next day, the protein was applied to a Ni^{2+}-IDA reverse-binding column. **Ni^{2+}-IDA Rebinding column (Column 2):** To a 25ml Econo-Pac polypropylene column (BioRad), 2ml of packed Ni-Sepharose resin was added, resin was washed in 3 column volumes of distilled deionized water then equilibrated in 5 column volumes of gel filtration buffer. The overnight protein plus TEV sample was loaded and passed through the column by gravity flow. The flow through was collected, to wash any remaining unbound protein from the column a 2x wash step of 2 column volumes of gel filtration buffer was included. Any bound protein was eluted with 2 column volume washes of 40mM and 500mM imidazole elution buffer. Collected fractions were analysed for verification of cleaved protein and purity by SDS-PAGE gel and mass spectrometry characterisation.
Step 3: SEC: Gel filtration	**Size Exclusion Chromatography (SEC) (Column 3):** Verified protein from rebinding column 2 was concentrated to required volume for gel filtration (<5 ml), using an Amicon-Ultra 10,000 MWCO centrifugal concentrator. The GF column (Superdex S200 16/60 HiLoad, 120ml on an ÄKTA Express chromatography system) was equilibrated in gel filtration buffer. The concentrated protein sample was loaded onto the GF column for further purification; protein was fractionated on the column in GF buffer at 1ml/min and 1ml fractions were collected at the A280 peaks. Peak fractions were analysed by SDS-PAGE gel, selected fractions containing high protein concentration and purity were collected and pooled, eliminating any bound contaminating non-specific proteins.
Step 4: Protein storage	**Protein concentration mg/ml and storage:** Protein was concentrated using a 10,000 Amicon-Ultra MWCO centrifugal concentrator and final protein concentration was determined from the

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	absorbance at 280nm, measured using a nanodrop spectrometer (Nanodrop ND-1000, NanoDrop Technologies). Final protein concentration was measured at 5mg/ml with a final volume of 11ml. Protein was transferred into 200 µl aliquots, flash-frozen in liquid nitrogen and stored at -80°C.
Note on Biotinylation	**Proteins expressed to a very high level in E. coli may not be completely biotinylated. To improve the outcome, shorter incubations and higher concentrations of Biotin in the medium are helpful. In addition, complete biotinylation may be achieved by incubation with purified BirA, ATP and Biotin in vitro.**

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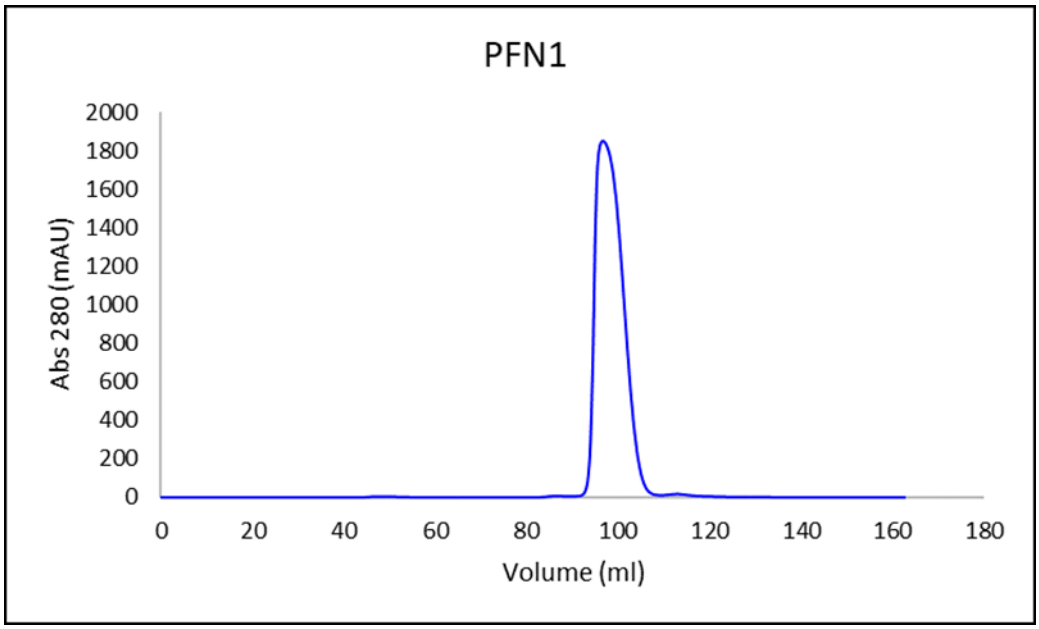
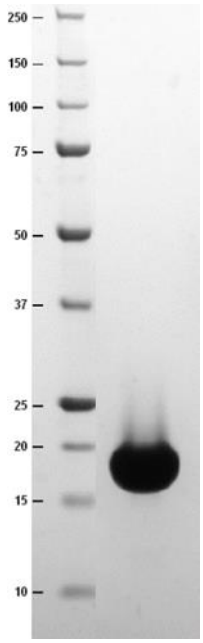
Gel Filtration Chromatogram (Superdex S200)



PFN1A-c000, Maximum peak elution time was 99.12ml with absorbance at 280nm of 1851.08 mAU. SEC analysis validates protein is a monomer

Final Sample SDS-PAGE



Protein	Expected mass + Biotinylation (Da)	Final concentration (mg/ml)
PFN1A-c000	17758	5

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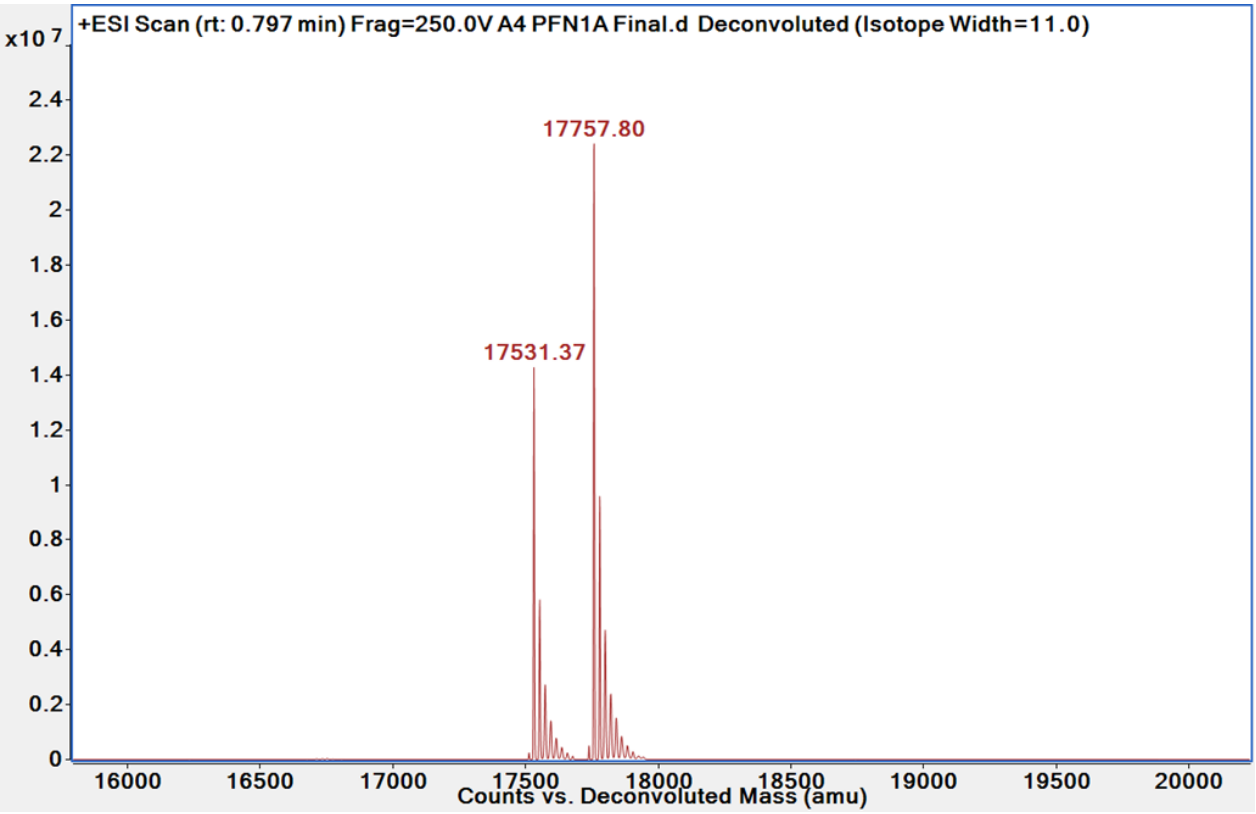
Mass Spectrometry characterisation

Protein	Vector	Expected mass cleaved + Biotinylation (Da)	Observed mass	Comments
PFN1A-c000	pNIC-Bio3	17758	17757.80 + 17531.37	60% Biotinylated 40% Non-Biotinylated



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